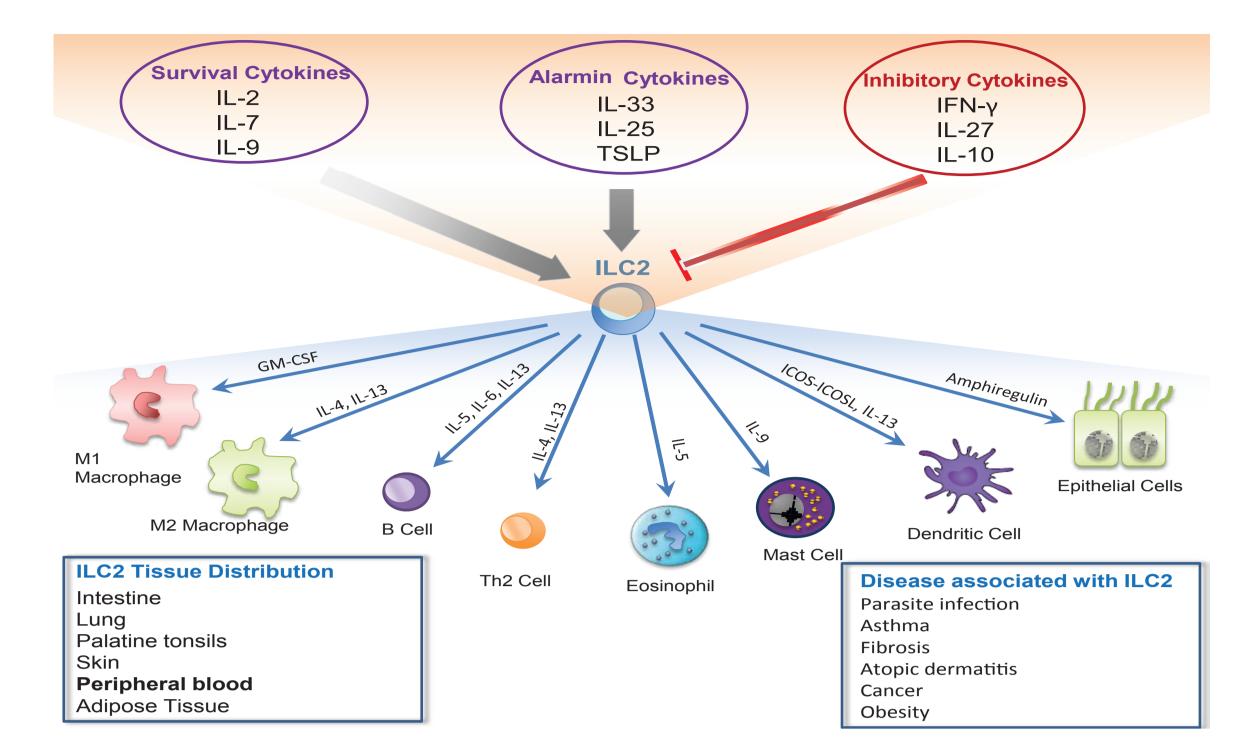
Fast and Efficient Enrichment of Functional ILC2 From Human Whole Blood

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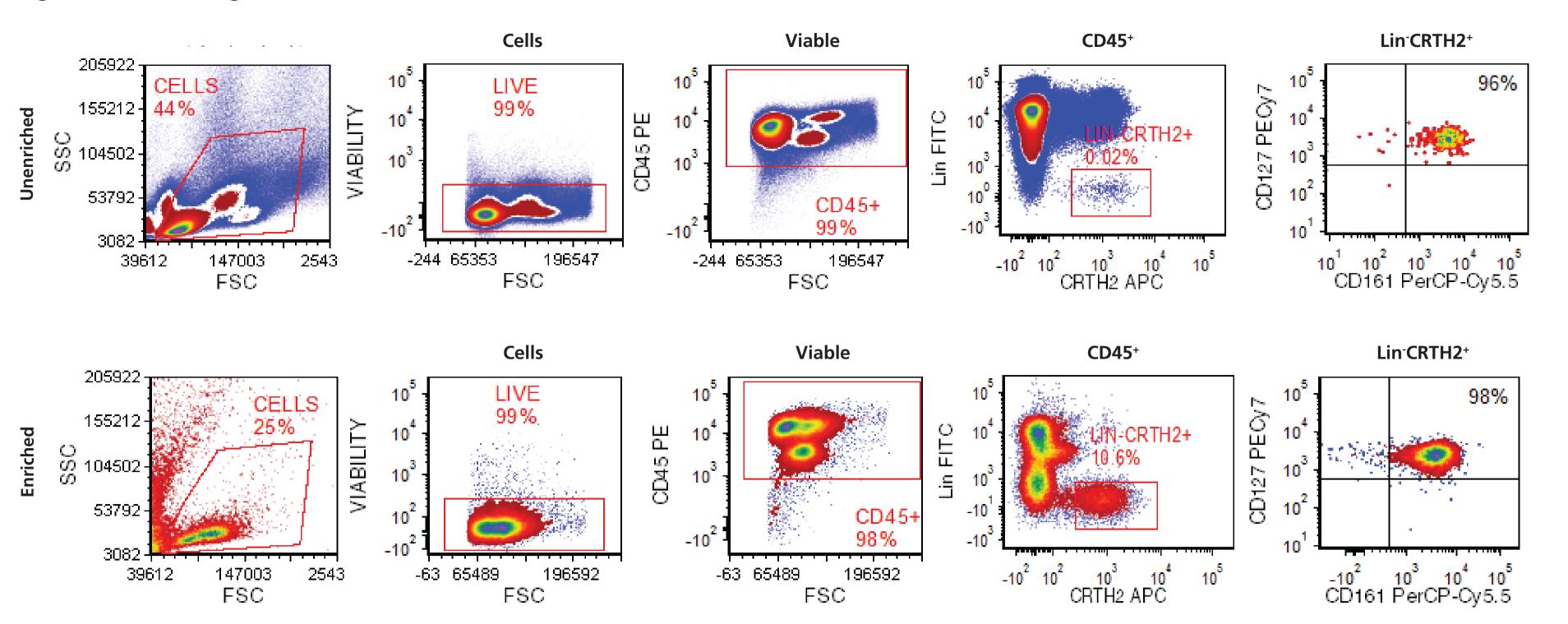
Introduction

Figure 1. Group-2 Innate Lymphoid Cells (ILC2)



Results ____

Figure 3. Percentage of ILC2 in whole blood, before and after enrichment



- Share characteristics with T cells and produce T helper cell associated cytokines
- Lack rearranged antigen-specific receptors
- Play a critical role in the pathology of Th-2 diseases such as allergies, asthma, and helminth infections, as well as metabolic diseases

ILC2 are difficult to study due to paucity, heterogeneity and similarity to other ILCs

- ILC2 are widely distributed throughout the body but they are extremely rare. The frequency of ILC2 in blood is approximately 0.01% of human leukocytes
- They do not express surface markers of mature lymphocytes and myeloid markers (Lin⁻). No unique markers specific for individual ILC populations are known
- Currently, multicolor flow cytometric cell sorting is the only method to isolate ILC2. However, sorting of ILC2 is time-consuming, expensive and often results in low purities and recoveries due to the scarcity of these cells in primary tissues
- GOAL: To develop a fast and simple procedure to pre-enrich ILC2 from whole blood by negative selection prior to cell sorting

Materials and Methods.

Samples

ILC2 were isolated either by density gradient centrifugation using Lymphoprep[™] (Cat# 07801) and SepMate[™] (Cat# 15450) (unenriched) or using RosetteSep[™] (enriched) as shown in Figure 2.

Gating strategy for human ILC2. Un-enriched and enriched samples were gated on WBC, LIVE, CD45⁺, Lin-CRTH2⁺, CD127⁺CD161⁺ cells. The gated population represents the percentage of ILC2 in whole blood before and after enrichment.

Figure 4. Percentage of ILC2, fold enrichment and number of ILC2 in blood before and after enrichment (n = 38)

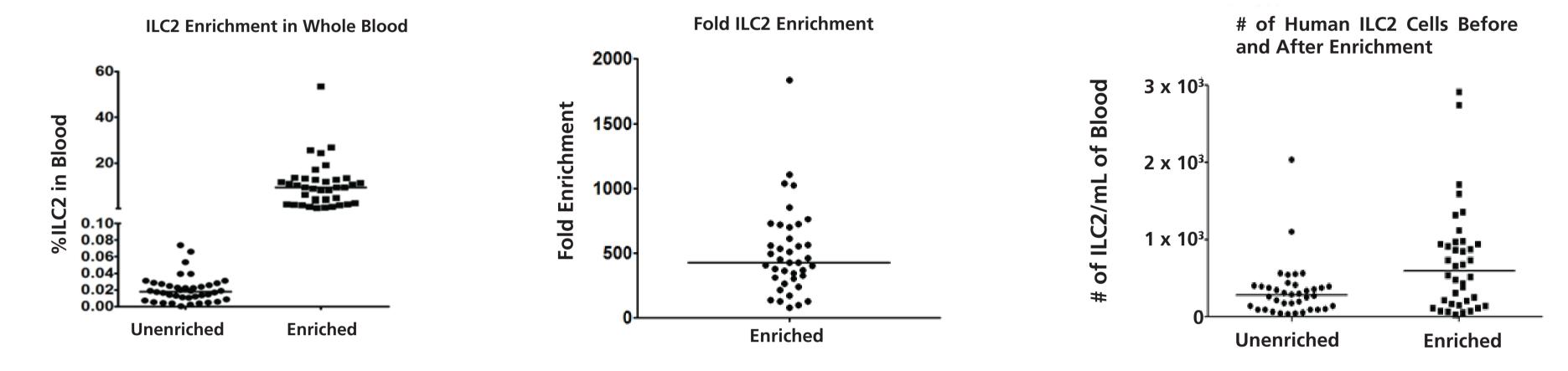


Table 1. Enrichment saves time and provides better ILC2 purity

Scenario 1: Focused on Recovery

Scenario 1	Unenriched Sample	RosetteSep™ Enriched Sample
Volume of whole blood processed and time	60 mL ammonium chloride lysis, 30 min	60 mL RosetteSep™ Enriched, 30 min
% of ILC2	0.01%	3%

Scenario 2: Focused on Purity

Scenario 2	Unenriched Sample	RosetteSep [™] - Enriched Sample
Volume of whole blood processed and time	60 mL (2X ammonium chloride lysis, 30 min)	60 mL RosetteSep [™] - Enriched, 30 min
% ILC2	0.05%	37%
Total Number of Cells Stained	200 x 10 ⁶	0.39 x 10 ⁶

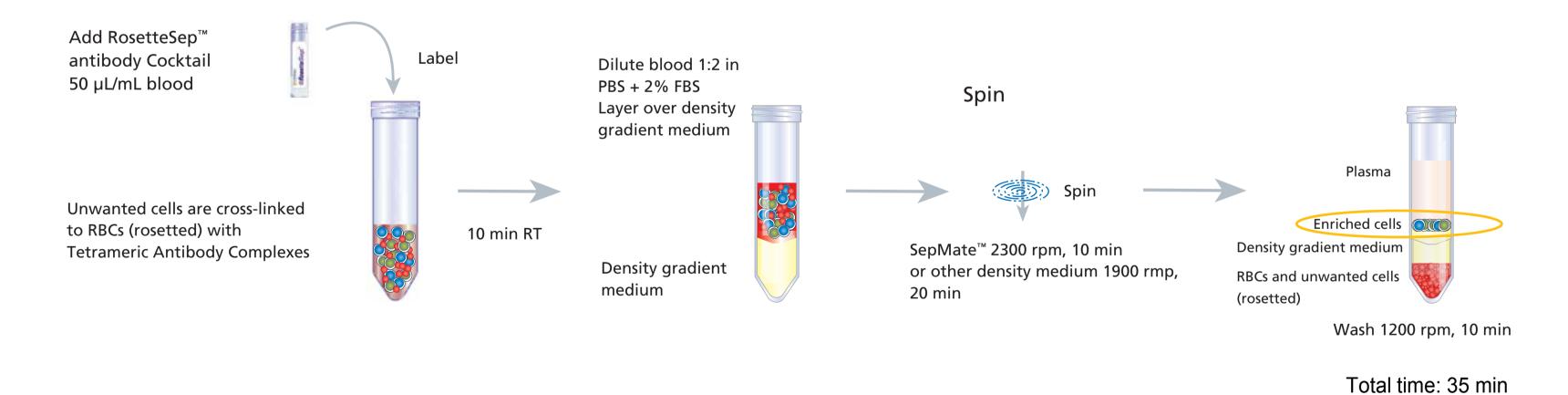
60 min

5 min

First Sort: Time

controls.





	Total Number of Cells Stained	130 x 10 ⁶	0.42 x 10 ⁶		
	First Sort: Time	87 min	8 min		
	Volume Sorted	60 mL (all processed)	All obtained after enrichment		
	Number of Cells	17489	10831		
	Purity	67%*	99%		
	Calculated # ILC2s post-sort	11738	10831		
*Second sort needed for good purity					

Calculated time to sort entire		
sample and # ILC obrained		

10% of stained All **Volume Sorted** sample Number of ILC2 3231 21305 Purity 46.75% 98.43% No second **Second Sort Time** 8 min sort Purity 97.16% 98.43% 21305 1719 Number of ILC2 ~ 11 hours 5 min ~ 17000 ILC2 21305

RosetteSep[™]-enriched cells were sorted and stimulated

using a cytokine cocktail. Pictures show proliferation at

different time points. Non-stimulated cells served as

Figure 5. RosetteSep[™]-enriched and sorted ILC2 are able to proliferate

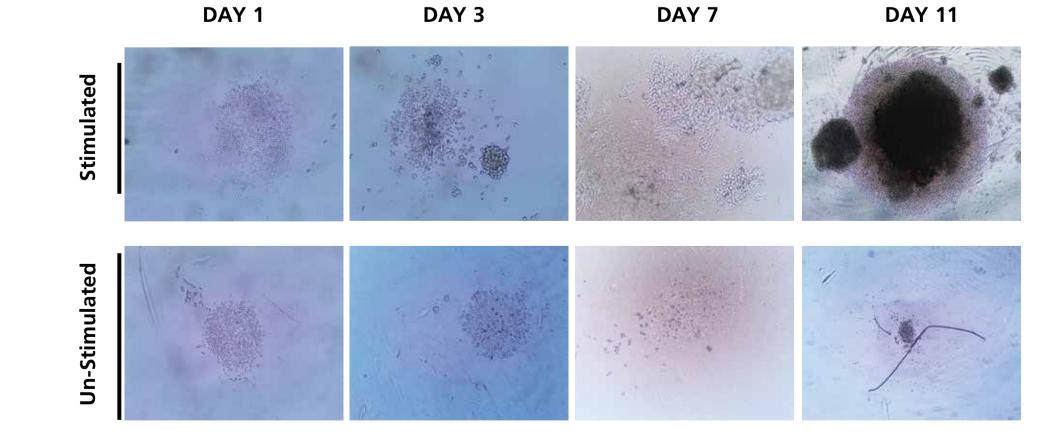


Figure 6. RosetteSep[™]-enriched ILC2s are functional and secrete IL-13

(A) IL-13 secretion by unenriched sorted human ILC2

IL-13 secretion by enriched sorted human ILC2

Cell Sorting

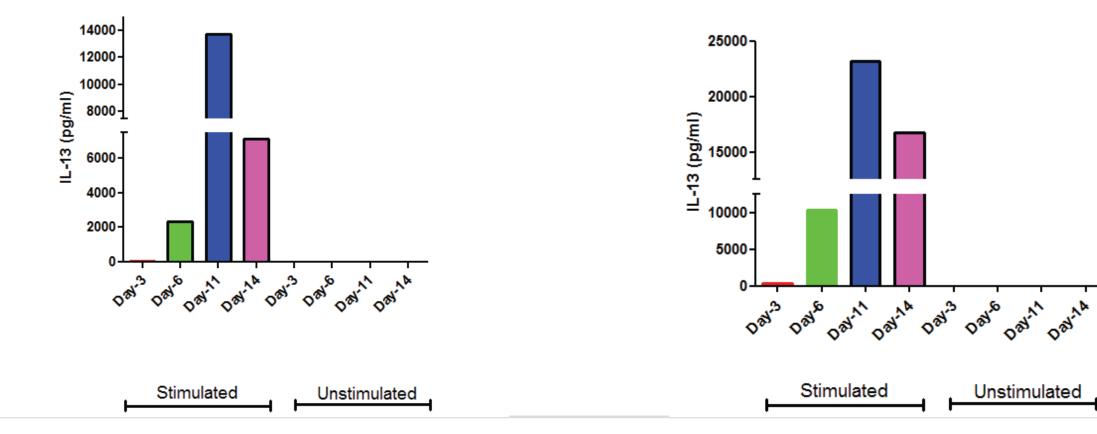
60 mL of unenriched or RosetteSep[™]-enriched samples were FACS sorted. Sorting time, purity and recovery were recorded and compared between the 2 approaches. Cells were cultured, stimulated and IL-13 production was analyzed by ELISA.

Assessment of ILC2

ILC2 were assessed using the following gating scheme: WBC \rightarrow LIVE (Viable) \rightarrow CD45⁺ \rightarrow LIN⁻CRTH2⁺ \rightarrow CD127⁺CD161⁺ Antibodies used in the lineage cocktail include, CD1a, CD3, CD11c, CD14, CD19, CD34, CD123, TCR $\alpha\beta$, TCR $\gamma\delta$, BDCA2, Fc ϵ R1

Conclusions.

- ILC2 frequency was between 0.002 to 0.07% after density gradient centrifugation alone (n=38)
- ILC2 were enriched to 0.44 53% with RosetteSep[™] (n=38)
- FACS sorting from enriched samples was faster (n=4, p<0.05 paired t test), and yielded higher ILC2 purity than FACS sorting from unenriched samples
- When stimulated in culture, sorted ILC2 from both enriched and unenriched samples secreted similar high levels of IL-13 as assessed by ELISA
- Enrichment of ILC2 prior to FACS sorting increased purity, shortened sorting time, and maintained ILC2 functionality



PBMCs were obtained by density gradient centrifugation (A) or RosetteSep[™]enrichment (B) prior to sorting for ILC2. Sorted cells were stimulated at different days and the supernatant analyzed for IL-13 by ELISA.



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