

**Negative Selection** 

Catalog #19851

For processing 1 x 10<sup>9</sup> cells



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TOLL FREE PHONE 1 800 667 0322 • PHONE +1 604 877 0713
INFO@STEMCELL.COM • TECHSUPPORT@STEMCELL.COM
FOR GLOBAL CONTACT DETAILS VISIT OUR WEBSITE

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## Description

Isolate untouched and highly purified T cells from mouse splenocytes by immunomagnetic negative selection. When using single-cell suspensions from other tissue types, this kit may require optimization.

- · Fast and easy-to-use
- · Up to 99% purity
- No column required
- · Untouched, viable cells

This kit targets non-T cells for removal with biotinylated antibodies recognizing specific cell surface markers. Unwanted cells are labeled with biotinylated antibodies and streptavidin-coated magnetic particles, and separated without columns using an EasySep<sup>TM</sup> magnet. Desired cells are simply poured off into a new tube. Isolated cells are immediately available for downstream applications such as flow cytometry, culture, and cell-based experiments.

# Component Descriptions

COMPONENT NAME	COMPONENT #	QUANTITY	STORAGE	SHELF LIFE	FORMAT
EasySep™ Mouse T Cell Isolation Cocktail	19851C.1	1 x 0.5 mL	Store at 2 - 8°C. Do not freeze.	Stable until expiry date (EXP) on label.	A combination of monoclonal antibodies in PBS and 0.1% BSA.
EasySep™ Streptavidin RapidSpheres™ 50001	50001	1 x 1 mL	Store at 2 - 8°C. Do not freeze.	Stable until expiry date (EXP) on label.	A suspension of magnetic particles in PBS.
Normal Rat Serum	13551	1 x 2 mL	Store at -20°C.	Stable until expiry date (EXP) on label.	Mycoplasma-free normal rat serum.

BSA - bovine serum albumin; PBS - phosphate-buffered saline

Components may be shipped at room temperature (15 - 25°C) but should be stored as indicated above.

## Additional Reagent Stability Information

REAGENT NAME	STORAGE	SHELF LIFE
Normal Rat Serum (in-use)	Store at 2 - 8°C.	Stable for at least 2 months. Do not exceed expiry date (EXP) on label.

## Sample Preparation

SPLEEN

Disrupt spleen in PBS or Hanks' Balanced Salt Solution (HBSS) containing 2% fetal bovine serum (FBS). Remove aggregates and debris by passing cell suspension through a 70 µm mesh nylon strainer. Centrifuge at 300 x g for 10 minutes and resuspend at 1 x 10^8 nucleated cells/mL in recommended medium. Ammonium chloride treatment is not recommended when preparing the cells for separation.

#### Recommended Medium

EasySep™ Buffer (Catalog #20144), RoboSep™ Buffer (Catalog #20104), or PBS containing 2% FBS and 1 mM EDTA. HBSS, Modified (Without Ca++ and Mg++; Catalog #37250) can be used in place of PBS. Medium should be free of Ca++, Mg++, and biotin.





# Directions for Use – Manual EasySep™ Protocols

See page 1 for Sample Preparation and Recommended Medium. Refer to Tables 1 and 2 for detailed instructions regarding the EasySep™ procedure for each magnet.

#### Table 1. EasySep™ Mouse T Cell Isolation Kit Protocol

_		EASYSEP™ MAGNETS	
STEP	INSTRUCTIONS	EasySep™ (Catalog #18000)	"The Big Easy" (Catalog #18001)
1	Prepare sample at the indicated cell concentration within the volume range.	1 x 10^8 cells/mL 0.1 - 2 mL	1 x 10^8 cells/mL 0.25 - 8 mL
2	Add Rat Serum to sample.	50 μL/mL of sample	50 μL/mL of sample
3	Add sample to required tube.	5 mL (12 x 75 mm) polystyrene round-bottom tube (e.g. Corning Catalog #352058)	14 mL (17 x 100 mm) polystyrene round-bottom tube (e.g. Corning Catalog #352057)
4	Add Isolation Cocktail to sample.	50 μL/mL of sample	50 μL/mL of sample
4	Mix and incubate.	RT for 10 minutes	RT for 10 minutes
5	Vortex RapidSpheres™. NOTE: Particles should appear evenly dispersed.	30 seconds	30 seconds
6	Add RapidSpheres™ to sample.	75 μL/mL of sample	75 μL/mL of sample
0	Mix and incubate.	RT for 2.5 minutes	RT for 2.5 minutes
7	Add recommended medium to top up the sample to the indicated volume. Mix by gently pipetting up and down 2 - 3 times.	Top up to 2.5 mL	<ul> <li>Top up to 5 mL for samples &lt; 4 mL</li> <li>Top up to 10 mL for samples ≥ 4 mL</li> </ul>
	Place the tube (without lid) into the magnet and incubate.	RT for 2.5 minutes	RT for 2.5 minutes
8	Pick up the magnet, and in one continuous motion invert the magnet and tube,* pouring the enriched cell suspension into a new tube.	Isolated cells are ready for use	Isolated cells are ready for use

RT - room temperature (15 - 25°C)

<sup>\*</sup> Leave the magnet and tube inverted for 2 - 3 seconds, then return upright. Do not shake or blot off any drops that may remain hanging from the mouth of the tube.





#### Table 2. EasySep™ Mouse T Cell Isolation Kit Protocol

		EASYSEPT	MAGNETS	
OTED	INSTRUCTIONS	EasyEights™ (Catalog #18103)		
STEP		5 mL tube	14 mL tube	
1	Prepare sample at the indicated cell concentration within the volume range.	1 x 10^8 cells/mL 0.25 - 2 mL	1 x 10^8 cells/mL 0.5 - 8 mL	
2	Add Rat Serum to sample.	50 μL/mL of sample	50 μL/mL of sample	
3	Add sample to required tube.	5 mL (12 x 75 mm) polystyrene round-bottom tube (e.g. Corning Catalog #352058)	14 mL (17 x 100 mm) polystyrene round-bottom tube (e.g. Corning Catalog #352057)	
	Add Isolation Cocktail to sample.	50 μL/mL of sample	50 μL/mL of sample	
4	Mix and incubate.	RT for 10 minutes	RT for 10 minutes	
5	Vortex RapidSpheres™. NOTE: Particles should appear evenly dispersed.	30 seconds	30 seconds	
	Add RapidSpheres™ to sample.	75 μL/mL of sample	75 μL/mL of sample	
6	Mix and incubate.	RT for 2.5 minutes	RT for 2.5 minutes	
7	Add recommended medium to top up the sample to the indicated volume. Mix by gently pipetting up and down 2 - 3 times.	Top up to 2.5 mL	<ul> <li>Top up to 5 mL for samples &lt; 4 mL</li> <li>Top up to 10 mL for samples ≥ 4 mL</li> </ul>	
	Place the tube (without lid) into the magnet and incubate.	RT for 5 minutes	RT for 5 minutes	
8	Carefully pipette (do not pour) the enriched cell suspension** into a new tube.	Isolated cells are ready for use	Isolated cells are ready for use	

RT - room temperature (15 - 25°C)

<sup>\*\*</sup> Collect the entire supernatant, all at once, into a single pipette (e.g. for the EasyEights™ 5 mL tube use a 2 mL serological pipette and for the EasyEights™ 14 mL tube use a 10 mL serological pipette).





# Directions for Use – Fully Automated RoboSep™ Protocol

See page 1 for Sample Preparation and Recommended Medium. Refer to Table 3 for detailed instructions regarding the RoboSep™ procedure.

#### Table 3. RoboSep™ Mouse T Cell Isolation Kit Protocol

STEP	INSTRUCTIONS	RoboSep™ (Catalog #20000 and #21000)	
1	Prepare sample at the indicated cell concentration within the volume range.	1 x 10^8 cells/mL 1 - 8 mL NOTE: If starting with 0.5 - 1 x 10^8 cells, resuspend cells in 1 mL.	
2	Add Rat Serum to sample.	50 μL/mL of sample	
3	Add sample to required tube.	14 mL (17 x 100 mm) polystyrene round-bottom tube (e.g. Corning Catalog #352057)	
4	Select protocol.	Mouse T Cell Isolation 19851	
5	Vortex RapidSpheres™. NOTE: Particles should appear evenly dispersed.	30 seconds	
6	Load the carousel.	Follow on-screen prompts	
6	Start the protocol.	Press the green "Run" button	
7	Unload the carousel when the run is complete.	Isolated cells are ready for use	

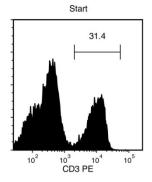
# Notes and Tips

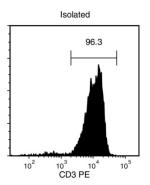
ASSESSING PURITY

For purity assessment of T cells by flow cytometry use the following fluorochrome-conjugated antibody clone:

• Anti-Mouse CD3e Antibody, Clone 145-2C11 (Catalog #60015)

## Data





Starting with mouse splenocytes, the T cell content (CD3+) of the isolated fraction typically ranges from 91.7 - 98.6%. In the above example, the purities of the start and final isolated fractions are 31.4% and 96.3%, respectively.

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