

## INTENDED USE

SepMate™ tubes are designed for the in vitro isolation of mononuclear cells (MNCs) from human whole peripheral blood and cord blood samples by density gradient centrifugation.

## PRODUCT DESCRIPTION

Polypropylene tube with insert. Gamma-irradiated.

## STORAGE

Store at room temperature (15 - 25°C).

## DIRECTIONS FOR USE

Ensure that sample, phosphate-buffered saline with 2% fetal bovine serum (PBS + 2% FBS; Catalog #07905), density gradient medium (see Notes on reverse page), and centrifuge are all at room temperature (15 - 25°C).

1. Add density gradient medium to the SepMate™ tube by carefully pipetting it through the central hole of the SepMate™ insert. Refer to Table 1 for required volumes. The top of the density gradient medium will be above the insert.

*Note: Small bubbles may be present in the density gradient medium after pipetting. This will not affect performance.*

2. Dilute sample with an equal volume of PBS + 2% FBS. Mix gently.

*For example, dilute 5 mL of sample with 5 mL of PBS + 2% FBS.*

3. Keeping the SepMate™ tube vertical, add the diluted sample by pipetting it down the side of the tube. The sample will mix with the density gradient medium above the insert.

*Note: The sample can be poured down the side of the tube. Take care not to pour the diluted sample directly through the central hole.*

4. Centrifuge at **1200 x g** (see Notes) for **10 minutes** at room temperature, with the **brake on**.

*Note: For samples older than 24 hours, a centrifugation time of 20 minutes is recommended.*

5. Pour off the top layer, which contains the enriched MNCs, into a new tube. Do not hold the SepMate™ tube in the inverted position for longer than 2 seconds.

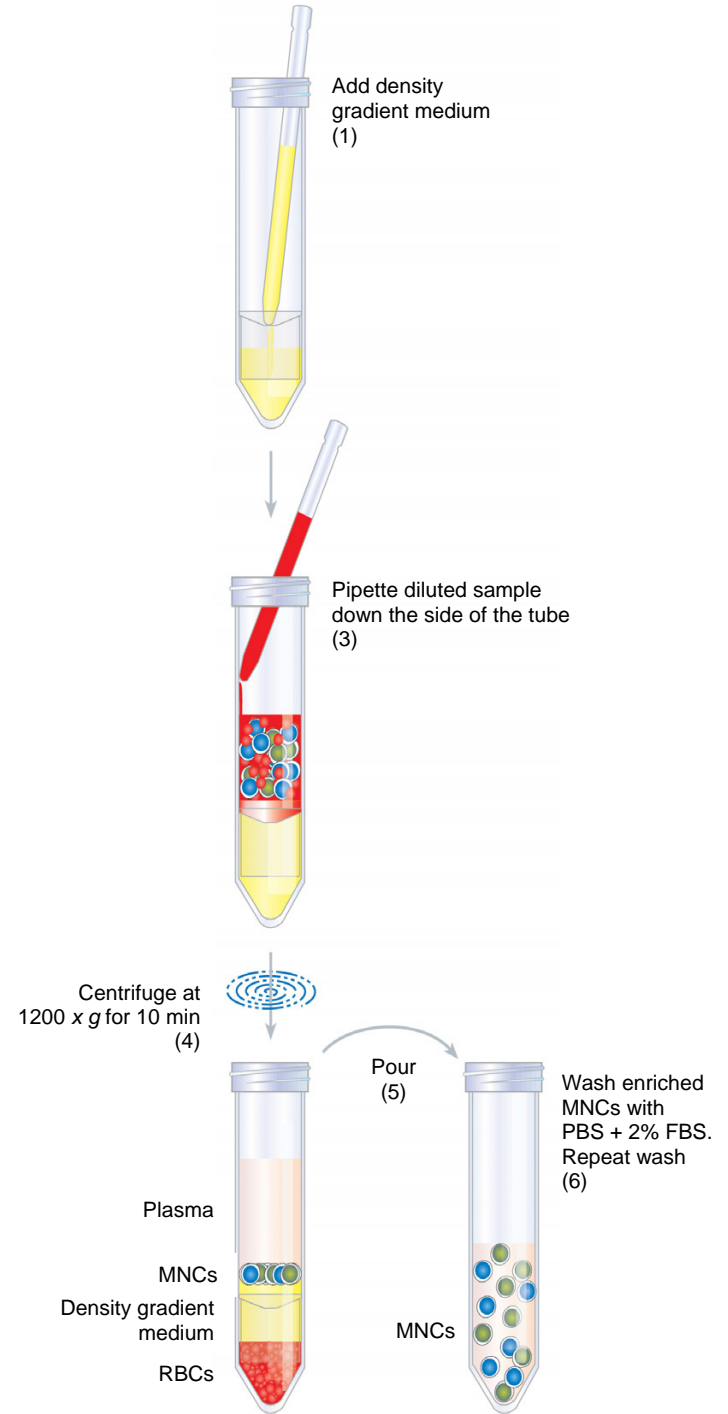
*Note: Some red blood cells (RBCs) may be present on the surface of the SepMate™ insert after centrifugation. This will not affect performance.*

6. Wash enriched MNCs with PBS + 2% FBS. Repeat wash.

*Note: Centrifuging at 300 x g for 8 minutes at room temperature, with the brake on, is recommended.*

## SEPMATE™ PROCEDURE

Numbers in brackets refer to steps under Directions for Use.



**Table 1: Density Gradient Medium Volumes**

SEPMATE™ TUBE	INITIAL BLOOD SAMPLE (mL)	DENSITY GRADIENT MEDIUM (mL)
15	0.5 - 4.0	4.5
15	> 4 - 5	3.5
50	4 - 17	15

FOR RESEARCH USE ONLY. NOT INTENDED FOR HUMAN OR ANIMAL DIAGNOSTIC OR THERAPEUTIC USES.

## NOTES

### Samples

SepMate™ can be used with human whole peripheral blood and cord blood samples. It has not been tested with samples older than 48 hours. For use of SepMate™ with samples other than human whole peripheral blood or cord blood please contact STEMCELL Technologies' Technical Support at techsupport@stemcell.com.

### SepMate™-15

SepMate™-15 is designed to process 0.5 - 5 mL of initial sample.

A minimum packed RBC volume of 0.25 mL is required. For patient samples with low hematocrits, the minimum sample volume may therefore be greater than 0.5 mL.

There is a maximum packed RBC volume of 3 mL. For patient samples with very high hematocrits, the maximum sample volume may therefore be less than 5 mL.

### SepMate™-50

SepMate™-50 is designed to process 4 - 17 mL of initial sample.

A minimum packed RBC volume of 2 mL is required. For patient samples with low hematocrits, the minimum sample volume may therefore be greater than 4 mL.

There is a maximum packed RBC volume of 12 mL. For patient samples with very high hematocrits, the maximum sample volume may therefore be less than 17 mL.

### Density Gradient Medium

Density gradient medium refers to Lymphoprep™ (Catalog #07801), Ficoll-Paque™ PLUS or other similar density gradient media.

### Recommended Medium

The recommended medium is phosphate-buffered saline with 2% fetal bovine serum (PBS + 2% FBS, Catalog #07905).

### Conversion of g to RPM

To convert g to rpm, use the following formula:

$$\text{RPM} = \sqrt{\frac{\text{RCF}}{(1.118 \times 10^{-5}) \times (\text{Radius})}}$$

Where: RPM = centrifuge speed in revolutions per minute

RCF = relative centrifugal force (g)

Radius = radius of centrifuge rotor in centimeters (cm)

### Troubleshooting

If the density gradient medium above the SepMate™ insert appears red after centrifugation (i.e. some RBCs have not pelleted), the SepMate™ tube can be spun at 1200 x g for another 10 minutes with the brake on. This situation may occur with samples that are older than 24 hours.

#### Platelet Removal (optional)

Platelets present in the plasma layer may be removed from the enriched MNCs in one of the following ways:

- In step 5, pipette off the supernatant above the MNC layer before pouring
- In step 6, perform one of the washes at 120 x g for 10 minutes at room temperature, with the brake off

## SUPPLEMENTARY PROCEDURE

### Use of SepMate™ with RosetteSep™ Cocktails

SepMate™ tubes can be used with RosetteSep™ cell enrichment cocktails to isolate specific cell types from human whole blood. For available RosetteSep™ cocktails please refer to [www.rosettesep.com](http://www.rosettesep.com).

To use SepMate™ with RosetteSep™ cocktails:

1. Add RosetteSep™ cocktail to the whole blood sample using volumes recommended in the RosetteSep™ cocktail Product Information Sheet.
2. Incubate for **10 minutes** at room temperature (15 - 25°C).  
*Note: The 10-minute incubation time is specific to this procedure. It will have minimal effect on performance.*
3. Follow the steps under SepMate™ Directions for Use, on reverse page.

*Note: Use density gradient medium recommended in the RosetteSep™ cocktail Product Information Sheet.*