

DIRECTIONS FOR USE

Ensure that blood sample, recommended medium, density gradient medium (see Notes and Tips, reverse page), and centrifuge are all at room temperature (15 - 25°C).

- Add EDTA to whole blood to a final concentration of 1 mM (e.g. add 10 µL of a 100 mM solution of EDTA per 1 mL of whole blood).
- Add RosetteSep™ Enrichment Cocktail at 50 µL/mL of whole blood (e.g. for 2 mL of whole blood, add 100 µL of cocktail). Mix well.
 Note: If using samples other than fresh whole blood, please see Notes and Tips.
- 3. Incubate 20 minutes at room temperature (15 25°C).
- Dilute sample with an equal volume of recommended medium and mix gently.
- Layer the diluted sample on top of the density gradient medium OR

Layer the density gradient medium underneath the diluted sample.

Note: Be careful to minimize mixing of the density gradient medium and sample.

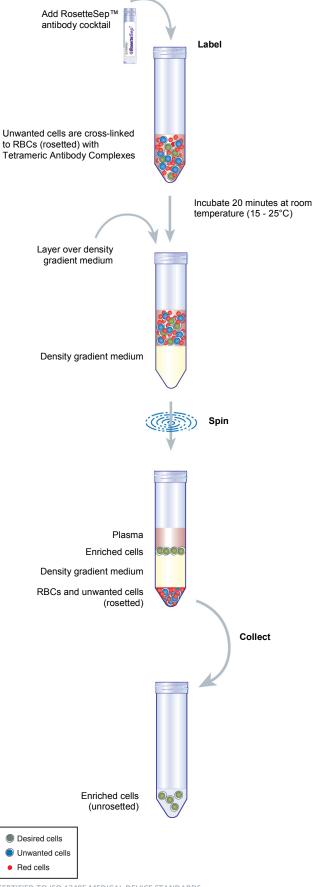
See table below for volume recommendations. With 50 mL centrifuge tubes, we suggest using a minimum of 15 mL density gradient medium to make it easier to remove the enriched cell layer.

WHOLE BLOOD (mL)	PBS + 2% FBS (mL)	DENSITY GRADIENT MEDIUM (mL)	TUBE SIZE (mL)
1	1	1.5	5
2	2	3	14
3	3	3	14
4	4	4	14
5	5	15	50
10	10	15	50
15	15	15	50

- Centrifuge for 20 minutes at 1200 x g (see Notes and Tips) at room temperature (15 - 25°C), with the brake off.
- Remove the enriched cells from the density gradient medium:plasma interface.

Note: Sometimes it is difficult to see the cells at the interface, especially when very rare cells are enriched. It is advisable to remove some of the density gradient medium along with the enriched cells in order to ensure their complete recovery.

- 8. Wash enriched cells with recommended medium. Repeat.
- Use enriched cells as desired. We recommend that enriched samples are lysed with Ammonium Chloride Solution (Catalog #07800) to remove residual red blood cells (RBCs) prior to flow cytometric analysis (this can be done as one of the wash steps) or if residual RBCs will interfere with subsequent assays.



ROSETTESEP™ PROTOCOL DIAGRAM

STEMCELL TECHNOLOGIES INC.'S QUALITY MANAGEMENT SYSTEM IS CERTIFIED TO ISO 13485 MEDICAL DEVICE STANDARDS. FOR RESEARCH USE ONLY. NOT INTENDED FOR HUMAN OR ANIMAL DIAGNOSTIC OR THERAPEUTIC USES.



2 mL 10 mL For labeling 40 mL of whole blood For labeling 200 mL of whole blood

© Rosette Sep™ PRODUCT INFORMATION SHEET

PRODUCT DESCRIPTION AND APPLICATIONS:

The RosetteSep™ Human Monocyte Enrichment Cocktail is designed to enrich monocytes from whole blood.

ROSETTESEP™ LABELING OF HUMAN CELLS

The RosetteSep™ antibody cocktail crosslinks unwanted cells in human whole blood to multiple RBCs, forming immunorosettes (Figure 1). This increases the density of the unwanted (rosetted) cells, such that they pellet along with the free RBCs when centrifuged over a density gradient medium. Desired cells are never labeled with antibody and are easily collected as a highly enriched population at the interface between the plasma and the density gradient medium.

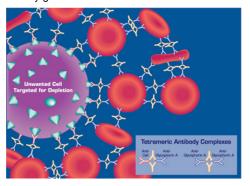


Figure 1 Rosette of unwanted cell and RBCs formed by RosetteSep™ Tetrameric Antibody Complexes (TACs)

NOTES AND TIPS

RECOMMENDED MEDIUM

The recommended medium is PBS containing 2% FBS and 1 mM EDTA (EasySep™ Buffer; Catalog #20144).

DENSITY GRADIENT MEDIUM

Density gradient medium refers to Lymphoprep™ (Catalog #07801), FicoII-Paque™ PLUS, or other similar density gradient media.

CONVERSION of g to RPM

To convert g to rpm, use the following formula:

RPM =
$$\sqrt{\frac{\text{RCF}}{(1.118 \times 10^{-5}) \times (\text{Radius})}}$$

Where: RPM = centrifuge speed in revolutions per minute

RCF = relative centrifugal force (g)

Radius = radius of centrifuge rotor in centimeters (cm)

SAMPLES OTHER THAN WHOLE BLOOD

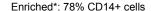
Although RosetteSep $^{\text{TM}}$ has been optimized for use with whole blood, cells can be enriched from other sources (i.e. buffy coat, leukaphereses). The concentration of nucleated cells in the sample should not exceed 5 x 10 7 cells/mL, and RBCs should be present at a ratio of at least 100 RBCs per nucleated cell.

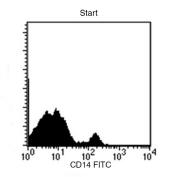
ASSESSING PURITY

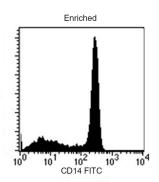
Purity of monocytes can be measured by flow cytometry after staining with a fluorochrome-conjugated anti-CD14 antibody such as Anti-Human CD14 Antibody, Clone M5E2 (Catalog #60004).

TYPICAL ROSETTESEP™ HUMAN MONOCYTE ENRICHMENT PROFILE:

Start*: 9.6% CD14+ cells







Starting with fresh peripheral blood the CD14+ cell content of the enriched fraction typically ranges from 72 - 85%.

*Note: Red blood cells were removed by lysis prior to flow cytometry.

COMPONENT DESCRIPTION:

ROSETTESEP™ HUMAN MONOCYTE ENRICHMENT COCKTAIL CODE #15028C.1

This cocktail contains a combination of monoclonal antibodies. These antibodies are bound in bispecific Tetrameric Antibody Complexes (TACs) which are directed against cell surface antigens on non-monocytes. It should be kept in mind that this product is a biological reagent, and as such cannot be completely characterized or quantified. Some variability is unavoidable.

Precipitate may be observed in the cocktail vial but will not affect performance.

STABILITY AND STORAGE:

ROSETTESEP™ HUMAN MONOCYTE ENRICHMENT COCKTAIL

Product stable at 2 - 8° C until expiry date as indicated on label. Do not freeze this product. This product may be shipped at room temperature (15 - 25° C), and should be refrigerated upon receipt.

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