BloodStor® 100

Biopreservation reagent for cells and tissues

<table>
<thead>
<tr>
<th>Catalog #</th>
<th>Volume</th>
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<tbody>
<tr>
<td>07951</td>
<td>50 mL</td>
</tr>
<tr>
<td>07939</td>
<td>100 mL</td>
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<tr>
<td>07938</td>
<td>5 x 100 mL</td>
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</tbody>
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Product Description

BloodStor® is a series of generic cGMP freezing media products used to cryopreserve stem cells and other cells isolated from umbilical cord blood, peripheral blood, bone marrow, and other biologics. BloodStor® 100 contains 100% dimethyl sulfoxide (DMSO) USP.

- Sterile, USP vial
- USP grade/highest-quality components
- Filled under cGMP
- Sterility and endotoxin testing

Properties

Storage: Store at 20 - 30°C.
Shelf Life: Stable until expiry date (EXP) on label. Protect product from prolonged exposure to light.
Contains: 100% Dimethyl sulfoxide (DMSO)

Please refer to the Safety Data Sheet (SDS) for hazard information.

BloodStor® 100 will solidify at temperatures below 18°C. If BloodStor® 100 is solidified upon receipt, allow to thaw at room temperature (18 - 25°C) and mix gently. Product performance will not be affected.

Handling / Directions For Use

FREEZING
1. Wipe down the outside of the cryopreservation reagent container and desired culture medium bottle with 70% ethanol or isopropanol before opening.
2. Prepare desired cryopreservation medium (e.g. 1 part BloodStor® 100 in 9 parts culture medium) and keep cold (2 - 8°C).
3. Obtain a cell suspension using a cell-specific protocol and centrifuge cells to obtain a cell pellet.
4. Carefully remove the supernatant with a pipette, leaving a small amount of medium to ensure the cell pellet is not disturbed. Resuspend the cell pellet by gently flicking the tube.
5. Add cold (2 - 8°C) cryopreservation medium, mix thoroughly, and transfer the suspension to a cryovial.
6. Freeze cells using a standard slow rate-controlled cooling protocol (approximately -1°C/minute) or an isopropanol freezing container and store at liquid nitrogen temperature (-135°C).

NOTE: Long-term storage at -80°C is not recommended.

THAWING
1. Warm medium of choice in a 37°C water bath.
2. Wipe the outside of the vial of cells with 70% ethanol or isopropanol.
3. In a biosafety hood, twist the cap a quarter-turn to relieve internal pressure and then retighten.
4. Quickly thaw cells in a 37°C water bath by gently shaking the vial. Remove the vial when a small frozen cell pellet remains. Do not vortex cells.
5. Wipe the outside of the vial with 70% ethanol or isopropanol.
6. Dilute in warmed medium of choice at a ratio of 1 part sample to 9 parts medium.
7. Centrifuge the cell suspension at 300 x g for 10 minutes at room temperature (15 - 25°C).
8. Carefully remove the supernatant with a pipette, leaving a small amount of medium to ensure the cell pellet is not disturbed. Resuspend the cell pellet by gently flicking the tube.
9. Gently add medium to the tube.
10. Repeat steps 7 and 8.