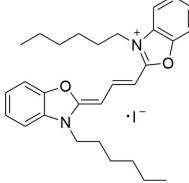
Dyes and		STENCELL TM
Stains	Green fluorescent lipophilic dye for staining cell membranes and other hydrophobic structures in	Scientists Helping Scientists [™] WWW.STEMCELL.CO
	live and fixed cells	TOLL FREE PHONE 1 800 667 0322 • PHONE +1 604 877 0
		INFO@STEMCELL.COM • TECHSUPPORT@STEMCELL.COM
Catalog #100-0815	25 mg	FOR GLOBAL CONTACT DETAILS VISIT OUR WEBSITE

Product Description

DiOC6(3) lodide, or 3,3-dihexyloxacarbocyanine iodide, is a green fluorescent lipophilic dye that is membrane permeable. At low concentrations, DiOC6(3) lodide accumulates in the mitochondria of live cells. At higher concentrations, DiOC6(3) lodide stains other membranes of live cells, including the endoplasmic reticulum and the Golgi apparatus. DiOC6(3) lodide also stains extracellular vesicles.

Chemical Name:	3,3'-dihexyloxacarbocyanine iodide	
Alternative Name:	3-hexyl-2-[3-(3-hexyl-2(3H)-benzoxazolylidene)-1-propen-1-yl]-benzoxazolium, iodide	
CAS Number:	53213-82-4	
Chemical Formula:	C ₂₉ H ₃₇ IN ₂ O ₂	
Molecular Weight:	572.52 g/mol	
Excitation Wavelength:	483 nm	
Emission Wavelength:	501 nm	
Structure:		



Properties

Storage: Shelf Life: Format: Store at -20°C. Stable until expiry date (EXP) on box label. Protect product from prolonged exposure to light. Red powder



Directions for Use

Please read the entire protocol before proceeding.

Preparation of DiOC6(3) Iodide Stock and Working Solutions

1. To prepare a stock solution, dissolve DiOC6(3) lodide in dimethyl sulfoxide (DMSO) or ethanol at 1 - 10 mM.

NOTE: If not used immediately, aliquot and store at -20°C. After thawing aliquots, use immediately; do not re-freeze.

 To prepare a DiOC6(3) lodide working solution, dilute the stock solution in a suitable buffer (e.g. phosphate-buffered saline [PBS]) to 1 - 10 μM. Use the working solution immediately; do not store.

NOTE: The optimal concentration of the working solution should be determined for different cell types and/or experimental conditions. For best results, test concentrations that span at least a 10-fold range.

Staining Cells

Refer to section A for staining cells in suspension or section B for staining adherent cells.

- A. CELLS IN SUSPENSION
- 1. Centrifuge cell suspension and remove and discard supernatant.
- 2. Resuspend cells at 1 x 10^6 cells/mL in DiOC6(3) lodide working solution in a conical tube (e.g. Catalog #38009).
- 3. Incubate at 37°C for 2 20 minutes; protect from light.

NOTE: The optimal incubation time should be determined for different cell types.

- 4. Centrifuge the cells at 110 250 x g for 5 minutes. Remove and discard supernatant.
- 5. Gently resuspend the cells in warm (37°C) culture medium to wash the cells.
- 6. Wash the cells 2 additional times by repeating steps 4 and 5.

B. ADHERENT CELLS

- 1. Culture adherent cells on a sterile glass coverslip until they reach the desired level of confluence.
- 2. Remove coverslip from culture medium and gently drain off excess medium. Place coverslip in a humidity chamber.
- 3. Pipette 100 µL of DiOC6(3) lodide working solution onto the corner of the coverslip and gently agitate until all cells are covered.
- 4. Incubate the coverslip at 37°C for 2 20 minutes; protect from light.

NOTE: The optimal incubation time should be determined for different cell types.

5. Drain off the solution and wash the coverslips 2 - 3X with culture medium. For each wash cycle, cover the cells with warm (37°C) culture medium, incubate at 37°C for 5 - 10 minutes protected from light, then drain off the medium.

Fluorescence Detection

- Microscopy: DiOC6(3) lodide can be detected with a standard FITC filter. 31001-Chroma and XF23-Omega optical filters are recommended.
- Flow cytometry: DiOC6(3) lodide-labeled cells can be analyzed using the conventional FL1 flow cytometer detection channel.

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