

# Human CD13 (ANPEP) ELISA Kit

For detection and measurement of human CD13

Catalog #100-0689

1 Kit



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## Product Description

The Human CD13 (ANPEP) ELISA Kit is designed for the quantitative detection and measurement of human CD13 in biological fluids such as serum, plasma, and tissue homogenates.

CD13, also known as aminopeptidase N or membrane alanyl aminopeptidase (ANPEP), is a type II membrane metalloprotease that consists of an extracellular-oriented catalytic domain. CD13 regulates the activity of numerous hormones, cytokines, and chemokines in inflammation. It is also coexpressed by MHC class II (MHC II)-bearing antigen-presenting cells (APCs), and is involved in the trimming of MHC II-associated peptides on the surface of APCs. It is an important marker for acute myeloid leukemia and plays a role in tumor invasion. CD13 is also known to be involved in the final digestion of peptides generated from the hydrolysis of proteins by gastric and pancreatic proteases. Research suggests that CD13 is also involved in viral infection. It regulates the human cytomegalovirus and is a receptor for human coronavirus 229E (HCoV-229E) Spike glycoprotein in HCoV-229E infections. It is also a potential therapeutic target for numerous inflammatory disorders, including rheumatoid arthritis.

The assay is based on the sandwich ELISA method, in which samples are added to ELISA strip plates pre-coated with capture antibodies specific for CD13. The captured CD13 is detected by the addition of a biotinylated detection antibody, followed by streptavidin-horseradish peroxidase (SA-HRP), which binds the biotinylated antibody. Addition of the chromogenic enzyme substrate 3,3',5,5' tetramethylbenzidine (TMB) results in a colored product with an intensity directly proportional to the concentration of human CD13. The concentration of CD13 is determined by comparison to a serial dilution of the standard analyzed in parallel.

## Product Information

All components listed below are stable until expiry date (EXP) on label. Kit may be shipped at room temperature (15 - 25°C) but should be stored at 2 - 8°C upon receipt. Once components are opened, store at 2 - 8°C and use within 1 month.

COMPONENT NAME	COMPONENT #	SIZE	STORAGE	DESCRIPTION
Human CD13 (ANPEP) Standard	300-0340	2 vials	Store at 2 - 8°C.	Lyophilized recombinant human CD13 (NS0-derived)
Human CD13 (ANPEP) ELISA Plate	300-0339	1 plate	Store at 2 - 8°C.	Plate (12 strips x 8 wells) coated with anti-human CD13 antibody
Human CD13 (ANPEP) Biotinylated Detection Antibody	300-0341	120 µL	Store at 2 - 8°C.	Biotinylated anti-human CD13 antibody (100X)
Human CD13 (ANPEP) Antibody Diluent	300-0343	15 mL	Store at 2 - 8°C.	For dilution of detection antibody
Adhesive Plate Covers	300-0349	4 covers	Store at 2 - 8°C.	For covering plates during incubation
Stop Solution*	300-0348	10 mL	Store at 2 - 8°C.	4 N Sulfuric acid
TMB Substrate	300-0347	10 mL	Store at 2 - 8°C.	3,3',5,5' tetramethylbenzidine (TMB) enzyme substrate solution
ELISA Diluent	300-0345	50 mL	Store at 2 - 8°C.	Buffer for dilution of standard and samples
SA-HRP	300-0342	120 µL	Store at 2 - 8°C.	Streptavidin-horseradish peroxidase (SA-HRP) conjugate
SA-HRP Diluent	300-0344	15 mL	Store at 2 - 8°C.	For dilution of SA-HRP
Wash Buffer (25X)	300-0346	20 mL	Store at 2 - 8°C.	Concentrated buffer solution for washing plates

\*Please refer to the Safety Data Sheet (SDS) for hazard information.

## Materials Required But Not Included

- Biohazard safety cabinet certified for Level II handling of biological materials
- Microplate reader set at a wavelength of 450 nm and (optional) a second correction wavelength set at 540 nm or 570 nm
- 37°C incubator
- ELISA plate washer: Automatic (adaptable for ELISA strip plates) or manual (e.g. multi-pipette or squirt bottle)
- Pipettor with appropriate tips
- Beakers, flasks, and graduated cylinders necessary for reagent preparations
- Tubes for standard and sample dilutions
- Timer
- Absorbent paper
- Deionized or distilled water

## Preparation of Reagents and Materials

### A. Wash Buffer

NOTE: Use clean or disposable glass or plasticware for preparation and storage of wash buffer.

NOTE: If crystals are visible in the concentrate, warm to room temperature and mix gently until completely dissolved.

Dilute Wash Buffer (25X) 1 in 25 with distilled or deionized water.

*Example: For one plate, prepare 500 mL wash buffer (1X) by adding 20 mL Wash Buffer (25X) to 480 mL distilled or deionized water.*

### B. ELISA Plate

1. Allow plate to adjust to room temperature (15 - 25°C) before opening the bags.
2. Plan the experiment to include a standard curve and background control (8 x 2 wells), a blank (2 wells), and test samples.
3. Assemble the required number of strips in the plate frame. Store the remaining strips in the foil bag containing the desiccant at 2 - 8°C for up to 1 month.

### C. Standard Stock Solution

NOTE: It is recommended to prepare fresh standard stock solution for each assay and use within 4 hours. However, reconstituted standard stock solution can be stored for 1 week at 2 - 8°C.

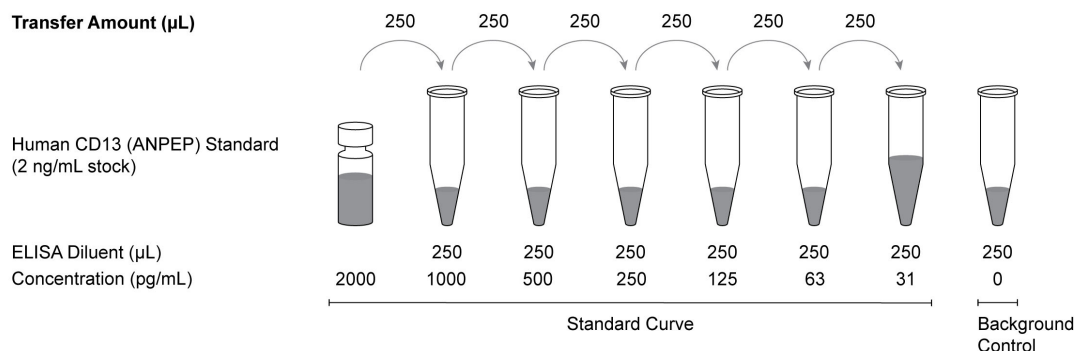
1. Centrifuge the standard vial to bring all material to the bottom of the vial.
2. Add 1 mL of ELISA Diluent to the vial of Human CD13 (ANPEP) Standard (final concentration 2000 pg/mL). Mix gently to ensure complete reconstitution (avoid foaming) and let sit for a minimum of 15 minutes.
3. Mix thoroughly.

### D. Standard Curve

NOTE: The standard curve dilutions should be prepared from standard stock solution and used within 1 hour. Duplicate wells for the standard curve dilutions and the background control are recommended.

Dilute the standard stock solution (prepared in section C) to create standard curve dilutions ranging from 31 - 2000 pg/mL according to Figure 1.

For the background control (0 pg/mL), use only ELISA Diluent.



NOTE: Volumes indicated are sufficient for duplicates.

**Figure 1. Recommended Serial Dilution of Standard**

## E. Samples

Refer to instructions below for preparing serum, plasma, or tissue samples. Avoid using lipemic, hemolyzed, or contaminated samples, as these may yield unreliable results. Fresh samples are recommended, to reduce protein degradation and denaturation risk. However, samples to be used within 5 days can be stored at 2 - 8°C. For samples to be used within 1 month, store at -20°C; for storage for 1 - 2 months, store samples at -80°C to avoid loss of bioactivity and contamination.

NOTE: Duplicate wells for all samples are recommended. For samples with a high protein concentration, it is advisable to prepare several dilutions for testing. Remove any particulate matter by centrifugation or filtration prior to use.

### Serum

1. Using a serum separator tube (SST), allow samples to clot at room temperature (15 - 25°C) for 2 hours or overnight at 2 - 8°C.
2. Centrifuge at 1000 x *g* for 15 minutes. Remove serum fraction and transfer to a fresh tube.
3. Dilute serum 1 in 100 in ELISA Diluent; this dilution may need to be optimized for individual samples. Samples containing high levels of protein (exceeding the range of the standard curve) will require further dilution.

NOTE: If not used immediately, aliquot and store at -20°C or -80°C. Avoid repeated freeze-thaw cycles.

### Plasma

1. Use EDTA or heparin as an anticoagulant when collecting plasma.
2. Within 30 minutes of collection, centrifuge plasma at 1000 x *g* for 15 minutes at 2 - 8°C. Remove and discard serum fraction, then transfer plasma to a fresh tube.
3. Dilute plasma 1 in 100 in ELISA Diluent; this dilution may need to be optimized for individual samples. Samples containing high levels of protein (exceeding the range of the standard curve) will require further dilution.

NOTE: If not used immediately, aliquot and store at -20°C or -80°C. Avoid repeated freeze-thaw cycles.

### Tissue homogenate

1. Collect 100 mg of tissue and rinse with phosphate-buffered saline (PBS).
2. Homogenize the tissue in 1 mL of PBS and store overnight at -20°C.
3. Perform two freeze-thaw cycles to break the cell membranes. Centrifuge the homogenate at 5000 x *g* for 5 minutes at 2 - 8°C.
4. Remove the supernatant and transfer to a fresh tube. If not used immediately, aliquot and store at -20°C or -80°C. Avoid repeated freeze-thaw cycles.

## F. Detection Antibody

1. Centrifuge the vial of Human CD13 (ANPEP) Biotinylated Detection Antibody to bring all material to the bottom of the vial.
2. Dilute Human CD13 (ANPEP) Biotinylated Detection Antibody 1 in 100 in Human CD13 (ANPEP) Antibody Diluent.

*Example: Add 10 µL of Human CD13 (ANPEP) Biotinylated Detection Antibody to 990 µL of Human CD13 (ANPEP) Antibody Diluent.*

## G. SA-HRP

Dilute SA-HRP 1 in 100 in SA-HRP Diluent.

*Example: Add 10 µL SA-HRP to 990 µL SA-HRP Diluent.*

## Directions for Use

Please read the entire protocol before proceeding. For each step in the protocol, total dispensing time for the addition of reagents or samples to the plate should not exceed 10 minutes. In all washing steps, each well must be thoroughly washed by filling with wash buffer, allowing to sit for 2 minutes, then blotting dry by tapping the plate upside down on absorbent paper.

NOTE: Cross-contamination of reagents may invalidate assay results. Permanently labeled, dedicated, multi-channel micropipette reservoirs for reagents are recommended.

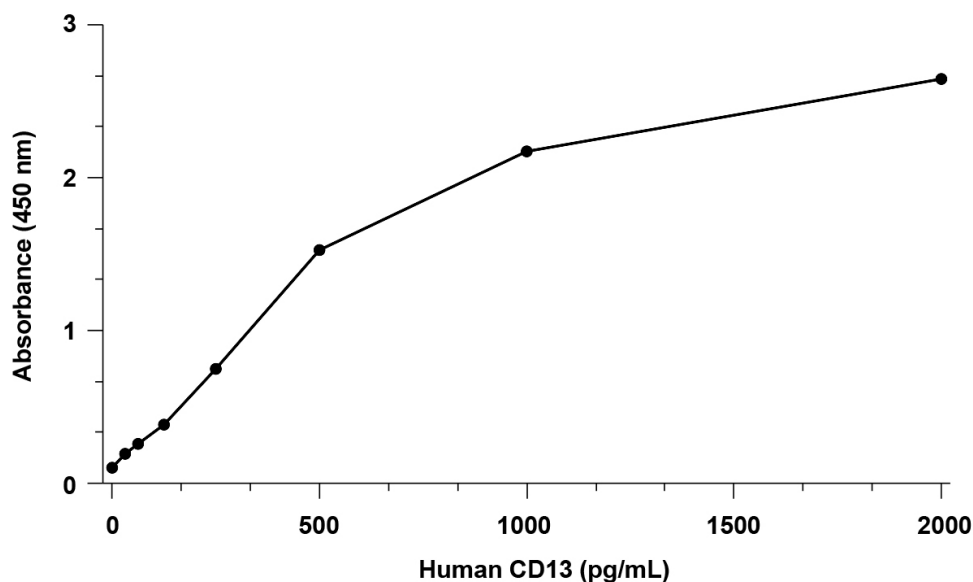
1. Centrifuge thawed samples to bring all material to the bottom of the vial. Bring all reagents and samples to room temperature (15 - 25°C), except the TMB Substrate, which should be kept at 2 - 8°C until use.
2. Prepare wash buffer (1X), reagents, samples, standard dilutions, and the ELISA plate as described in Preparation of Reagents and Materials.
3. Add 100 µL/well of each standard curve dilution and background control to ELISA plate. Leave the blank wells empty.
4. Add 100 µL/well of diluted test sample to ELISA plate.

NOTE: Duplicate wells for all samples are recommended.

5. Cover the plate with an Adhesive Plate Cover and incubate at 37°C for 2 hours.
6. Remove the liquid in each well and blot dry. **Do not wash the wells.**

7. Add 100  $\mu\text{L}$ /well of Detection Antibody. Leave blank wells empty.  
NOTE: If Detection Antibody appears cloudy, warm to room temperature and mix gently until solution appears uniform.
8. Cover the plate with an Adhesive Plate Cover and incubate at 37°C for 1 hour.
9. Wash each well with 5 x 200  $\mu\text{L}$  of wash buffer and blot dry.
10. Add 100  $\mu\text{L}$ /well of diluted SA-HRP. Leave blank wells empty.
11. Cover the plate with an Adhesive Plate Cover and incubate at 37°C for 1 hour.
12. Wash each well with 5 x 200  $\mu\text{L}$  of wash buffer and blot dry.
13. Add 90  $\mu\text{L}$ /well of TMB Substrate to all wells (including blank wells).  
NOTE: TMB Substrate should be colorless or light blue until added to the plate. Protect TMB Substrate from light.
14. Incubate at 37°C in the dark for 15 - 30 minutes.
15. Add 50  $\mu\text{L}$ /well of Stop Solution to all wells (including blank wells), in the same order as TMB Substrate addition. Gently tap the plate to ensure thorough mixing.  
NOTE: Use care when handling Stop Solution. Please refer to the Safety Data Sheet (SDS) for hazard information.  
NOTE: The color in the wells will turn from blue to yellow after addition of Stop Solution. Wells that are green indicate they are not thoroughly mixed.
16. Within 5 minutes of adding Stop Solution, measure the absorbance at 450 nm in a microplate reader. If possible, use a correction wavelength of 540 nm or 570 nm. Blank the reader using ELISA wells containing TMB Substrate and Stop Solution only.  
NOTE: Before measuring absorbance, remove any air bubbles in the wells using a small hypodermic needle or a pipette tip.
17. Calculate the average absorbance of each sample from the duplicate values. Subtract the mean absorbance value of the blank from the standard, the background control, and the sample values prior to generating the standard curve and determining the protein concentrations in the samples. A representative standard curve is shown in Figure 2.  
NOTE: Multiply the protein concentrations by the dilution factor used for each sample.

## Performance of the Assay



**Figure 2. Representative Standard Curve**

- **Reportable range:** 31.25 - 2000 pg/mL. This is the concentration range in which measurement of the analyte can be done with the highest precision, accuracy, and linearity.
- **Sensitivity:** The lower limit of detection (LLD) of this assay is 7.81 pg/mL. This is the lowest protein concentration that could be differentiated from zero as determined by the mean OD value of 20 replicates of the zero standard added by their three standard deviations.
- **Precision:** The intra-assay precision (CV) of this assay is < 8%. The inter-assay precision (CV) of this assay is < 10%.
- **Specificity:** This assay has high specificity for the detection of human CD13. No significant cross-reactivity or interference is known to occur between human CD13 and analogues.

## Troubleshooting

PROBLEM	POSSIBLE CAUSE	RECOMMENDED ACTION
Absorbance values too low	Incubation time too short	Follow the directions for the multiple incubation steps outlined in Directions for Use.
	Undiluted Wash Buffer (25X)	Dilute Wash Buffer (25X) with distilled or deionized water as described in Preparation of Reagents and Materials, section A.
	Incorrect dilution of Detection Antibody and/or SA-HRP	Follow the directions for dilution of Detection Antibody and SA-HRP in Preparation of Reagents and Materials.
	Wash step omitted before adding SA-HRP	Follow sequence of incubation and wash steps as outlined in Directions for Use.
Absorbance values too high	Incubation time too long	Follow the directions for the multiple incubation steps outlined in Directions for Use.
	Insufficient washing	Ensure that each well is washed 5X with 200 $\mu$ L of diluted Wash Buffer. Ensure that the multi-channel pipette fills and empties reproducibly. Blot wells dry before proceeding to the next step. If using an automated plate washer, add a 30-second soak period following the addition of wash buffer, and/or rotate the plate 180 degrees between wash steps.
	Incorrect dilution of Detection Antibody and/or SA-HRP	Follow the directions for dilution of Detection Antibody and SA-HRP in Preparation of Reagents and Materials.
Low absorbance readings despite good color development in wells	Incorrect wavelength setting on the ELISA reader	Check that the measuring wavelength is set at 450 nm and that the ELISA reader has the correct filter for this wavelength.
High background	Insufficient washing	Ensure that each well is washed 5X with 300 $\mu$ L of diluted Wash Buffer. Ensure that the multi-channel pipette fills and empties reproducibly. Blot wells dry before proceeding to the next step.
	Incorrect dilution of Detection Antibody and/or SA-HRP	Follow the directions for dilution of Detection Antibody and SA-HRP in Preparation of Reagents and Materials.
No signal with standard curve dilutions	Reagents not added in correct sequence	Follow sequence of incubation and wash steps as outlined in Directions for Use.
	Incorrect dilution of Detection Antibody and/or SA-HRP	Follow the directions for dilution of Detection Antibody and SA-HRP in Preparation of Reagents and Materials.
Inconsistent dose-response curve	Incubation conditions for individual wells not identical	Avoid delays during filling of assay wells; ensure that all reagents are at 15 - 25°C prior to beginning the assay.
Poor replicates	Insufficient washing	Ensure that each well is washed 5X with 200 $\mu$ L of diluted Wash Buffer. Ensure that the multi-channel pipette fills and empties reproducibly. Blot wells dry before proceeding to the next step. If using an automated plate washer, add a 30-second soak period following the addition of wash buffer, and/or rotate the plate 180 degrees between wash steps.
	Unequal volumes in wells	Ensure that pipettes function properly.
	Carry-over between wells; evaporation from wells; splashing of well contents onto adhesive cover	Always use a new adhesive cover for each incubation. Ensure that each well is sealed tightly. Keep filled plates in a horizontal position and handle with caution.
	Samples not mixed after thawing	Vortex samples after thawing.
	High concentration of lipids or particulate matter in samples	Filter or centrifuge samples to pellet aggregates. Lipids may concentrate on the surface after centrifugation and may be removed.
	Air bubbles in well during measurement with the ELISA reader	Remove air bubbles using a small hypodermic needle or pipette tip.
Absorbance values of sample dilutions decrease with increasing concentration	Protein concentration of the sample dilutions exceeds the upper limit of the assay	Dilute the samples further and retest along with the standard curve dilutions.
Unexpected ELISA results for tissue or cell extraction samples	Certain chemicals present in the chemical lysis buffer	Try different lysis buffers

## Related Products

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