## Directions for Use – Fully Automated RoboSep™ Protocol

See page 1 for Sample Preparation and Recommended Medium. Refer to Table 3 for detailed instructions regarding the RoboSep™ procedure.

**Table 3. RoboSep™ Human CD4+ T Cell Isolation Kit Protocol**

| STEP | INSTRUCTIONS | RoboSep™ (Catalog #20000 and #21000)  |
|---|---|---|
| 1 | Prepare sample at the indicated cell concentration within the volume range. | 5 x 10^7 cells/mL<br>0.5 - 8.5 mL |
| | Add sample to required tube. | 14 mL (17 x 95 mm) polystyrene round-bottom tube<br>(e.g. Catalog #38008) |
| 2 | Select protocol. | Human CD4+ T Cell Isolation 17952 |
| 3 | Vortex RapidSpheres™.<br>NOTE: Particles should appear evenly dispersed. | 30 seconds |
| 4 | Load the carousel. | Follow on-screen prompts |
| | Start the protocol. | Press the green “Run” button |
| 5 | Unload the carousel when the run is complete. Remove the tube containing the isolated cells. | Isolated cells are ready for use |

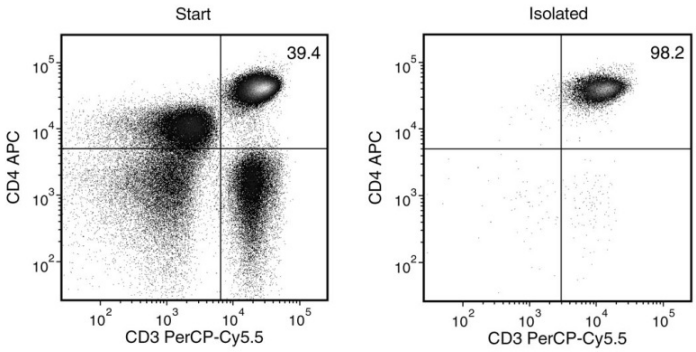
## Notes and Tips

ASSESSING PURITY

For purity assessment of CD4+ T cells (CD3+CD4+) by flow cytometry, use the following fluorochrome-conjugated antibodies:

- Anti-Human CD3 Antibody, Clone UCHT1 (Catalog #60011), and
- Anti-Human CD4 Antibody, Clone OKT4 (Catalog #60016)

## Data




Starting with human PBMCs, the CD4+ T cell content (CD3+CD4+) of the isolated fraction is typically 94.8 ± 2.3% (mean ± SD using the purple EasySep™ Magnet). In the above example, the purities of the start and final isolated fractions are 39.4% and 98.2%, respectively.