EasySep™ Human Naïve CD8+ T Cell Isolation Kit

For processing 1 x 10⁹ cells

Catalog #19258

Negative Selection

Document #10000005277 | Version 01



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Description

Isolate untouched and highly purified naïve CD8+ T cells (CD8+CD45RA+CCR7+ and CD45RO-CD57-CD56-) from fresh human peripheral blood mononuclear cells (PBMCs) by immunomagnetic negative selection.

- · Fast, easy-to-use and column-free
- Up to 97% purity
- Untouched, viable cells

This kit targets non-naïve CD8+ T cells for removal with antibodies recognizing specific cell surface markers. Unwanted cells are labeled with antibodies and magnetic particles, and separated without columns using an EasySep™ magnet. Desired cells are simply poured off into a new tube. Isolated cells are immediately available for downstream applications such as flow cytometry, culture, or DNA/RNA extraction.

Component Descriptions

COMPONENT NAME	COMPONENT #	QUANTITY	STORAGE	SHELF LIFE	FORMAT
EasySep Human Naïve CD8+ T Cell Isolation Cocktail	19258C	1 x 1 mL	Store at 2 - 8°C. Do not freeze.	Stable until expiry date (EXP) on label.	A combination of monoclonal antibodies in PBS.
EasySep™ D2 Magnetic Particles	19650	2 x 1 mL*	Store at 2 - 8°C. Do not freeze.	Stable until expiry date (EXP) on label.	A suspension of magnetic particles in TBS.

PBS - phosphate-buffered saline; TBS - TRIS-buffered saline

Components may be shipped at room temperature (15 - 25°C) but should be stored as indicated above.

Sample Preparation

For available fresh and frozen samples, see www.stemcell.com/primarycells.

PERIPHERAL BLOOD

Prepare a PBMC suspension from whole blood by centrifugation over a density gradient medium (e.g. Lymphoprep[™], Catalog #07801). For more rapid PBMC preparation, use the SepMate[™] RUO (Catalog #86450/86415) or SepMate[™] IVD* (Catalog #85450/85415) cell isolation tube.

After preparation, resuspend cells at 5 x 10^7 cells/mL in recommended medium.

* SepMate™ IVD is only available in select regions where it is registered as an In Vitro Diagnostic (IVD) device for the isolation of mononuclear cells (MNCs) from whole blood or bone marrow by density gradient centrifugation. In all other regions SepMate™ is available for research use only (RUO).

LEUKAPHERESIS

- Add an equal volume of Ammonium Chloride Solution (Catalog #07800) to the leukapheresis sample.
 NOTE: If working with large volumes (> 150 mL), concentrate leukapheresis sample first by centrifuging at 500 x g for 10 minutes. Remove the supernatant and resuspend the cells in 1/10th of the original leukapheresis volume with recommended medium (e.g. for 300 mL of cells, resuspend in 30 mL of recommended medium). For small volumes (≤ 150 mL), add Ammonium Chloride Solution directly to the leukapheresis sample.
- 2. Incubate on ice for 15 minutes.
- 3. Centrifuge at 500 x g for 10 minutes at room temperature (15 25°C). Remove the supernatant.
- 4. Wash the cells by topping up the tube with recommended medium. Centrifuge the cells at 150 x g for 10 minutes at room temperature with the brake off. Carefully remove the supernatant.
- 5. Repeat step 4 one or more times until most of the platelets have been removed (indicated by a clear supernatant).
- 6. Resuspend the cells at 5 x 10^7 cells/mL in recommended medium.

Recommended Medium

EasySep™ Buffer (Catalog #20144), RoboSep™ Buffer (Catalog #20104), or PBS containing 2% fetal bovine serum (FBS) and 1 mM EDTA. Medium should be free of Ca++ and Mg++.

^{*} If RoboSep™ Human Naïve CD8+ T Cell Isolation Kit (Catalog #19258RF) has been ordered, the kit will come with 5 x 1 mL of EasySep™ D2 Magnetic Particles.



Directions for Use – Manual EasySep™ Protocols

See page 1 for Sample Preparation and Recommended Medium. Refer to Table 1 for detailed instructions regarding the EasySep™ procedure for each magnet.

Table 1. EasySep Human Naïve CD8+ T Cell Isolation Kit Protocol

		EASYSEP™ MAGNETS			
STEP	INSTRUCTIONS	EasySep™ (Catalog #18000)	"The Big Easy" (Catalog #18001)		
1	Prepare sample at the indicated cell concentration within the volume range.	5 x 10^7 cells/mL 0.25 - 2 mL	5 x 10^7 cells/mL 0.5 - 8 mL		
	Add sample to required tube.	5 mL (12 x 75 mm) polystyrene round-bottom tube (e.g. Catalog #38007)	14 mL (17 x 95 mm) polystyrene round-bottom tube (e.g. Catalog #38008)		
2	Add Isolation Cocktail to sample.	50 μL/mL of sample	50 μL/mL of sample		
2	Mix and incubate.	RT for 10 minutes	RT for 10 minutes		
3	Vortex Magnetic Particles. NOTE: Particles should appear evenly dispersed.	30 seconds	30 seconds		
4	Add Magnetic Particles to sample.	100 μL/mL of sample	100 μL/mL of sample		
	Mix and incubate.	RT for 10 minutes	RT for 10 minutes		
5	Add recommended medium to top up the sample to the indicated volume. Mix by gently pipetting up and down 2 - 3 times.	Top up to 2.5 mL	 Top up to 5 mL for samples < 3 mL Top up to 10 mL for samples ≥ 3 mL 		
	Place the tube (without lid) into the magnet and incubate.	RT for 5 minutes	RT for 5 minutes		
6	Pick up the magnet, and in one continuous motion invert the magnet and tube,* pouring the enriched cell suspension into a new tube.	Use a new 5 mL tube	Use a new 14 mL tube		
7	Remove the tube from the magnet and place the new tube (without lid) into the magnet and incubate for a second separation.	RT for 5 minutes	RT for 5 minutes		
8	Pick up the magnet, and in one continuous motion invert the magnet and tube,* pouring the enriched cell suspension into a new tube.	Isolated cells are ready for use	Isolated cells are ready for use		

RT - room temperature (15 - 25°C)

^{*} Leave the magnet and tube inverted for 2 - 3 seconds, then return upright. Do not shake or blot off any drops that may remain hanging from the mouth of the tube.



Directions for Use – Fully Automated RoboSep™ Protocol

See page 1 for Sample Preparation and Recommended Medium. Refer to Table 3 for detailed instructions regarding the RoboSep™ procedure.

Table 3. RoboSep™ Human Naïve CD8+ T Cell Isolation Kit Protocol

STEP	INSTRUCTIONS	RoboSep™ (Catalog #20000 and #21000)	
1	Prepare sample at the indicated cell concentration within the volume range.	5 x 10^7 cells/mL 0.5 - 8 mL	
	Add sample to required tube.	14 mL (17 x 95 mm) polystyrene round-bottom tube (e.g. Catalog #38008)	
2	Select protocol.	For volumes between 0.5 - 2.9 mL: Human Naive CD8+ T Cell Isolation 19258 - small volume For volumes between 3 - 8 mL: Human Naive CD8+ T Cell Isolation 19258 - large volume	
3	Vortex Magnetic Particles. NOTE: Particles should appear evenly dispersed.	30 seconds	
	Load the carousel.*	Follow on-screen prompts	
4	Start the protocol.	Press the green "Run" button	
5	Unload the carousel when the run is complete.	Isolated cells are ready for use	

^{*} Two vials of EasySep™ D2 Magnetic Particles and two RoboSep™ Filter Tip Racks (Catalog #20125) may be required for a single run.

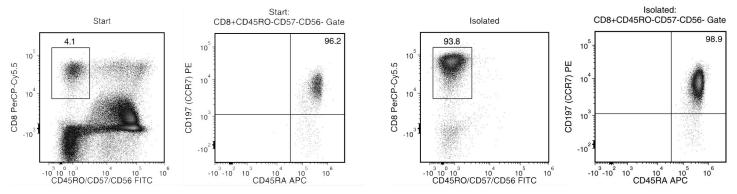
Notes and Tips

ASSESSING PURITY

For purity assessment of naïve CD8+ T cells (CD8+CD45RA+CCR7+ and CD45RO-CD57-CD56-) by flow cytometry, use the following fluorochrome-conjugated antibodies:

- · Anti-Human CD8a Antibody, Clone RPA-T8 (Catalog #60022), and
- Anti-human CD45RA antibody, and
- · Anti-human CD197 (CCR7) antibody, and
- · Anti-Human CD45RO Antibody, Clone UCHL1 (Catalog #60097), and
- · Anti-human CD57 antibody, and
- Anti-Human CD56 (NCAM) Antibody, Clone HCD56 (Catalog #60021)

Data



Starting with fresh PBMCs, the naïve CD8+ T cell content (CD8+CD45RA+CCR7+ and CD45RO-CD57-CD56-) of the isolated fraction is typically 92.3 ± 4.0% (mean ± SD using the purple EasySepTM Magnet). In the above example, the purities of the start and final isolated fractions are 3.9% and 92.8%, respectively.

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